Eileen J. Kennedy, Ph.D.

Dr. Samuel C. Benedict Professor
Interim Department Head
Director, UGA Cancer Center
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CURRENT POSITION

2022-Present	Co-Director, UGA Cancer Center
2022-Present	Interim Department Head, University of Georgia Department of Pharmaceutical and Biomedical Sciences
2019-Present	Dr. Samuel C. Benedict Professor, University of Georgia College of Pharmacy
2017-Present	Associate Prof. with Tenure, University of Georgia

EDUCATION

2006-2010	American Cancer Society Postdoctoral Fellow, Harvard University Advisor: Gregory L. Verdine, Chemistry and Chemical Biology
2000-2005	Ph.D. Chemistry, American Heart Association Predoctoral Fellow, UCSD Advisor: Gourisankar Ghosh, Chemistry and Biochemistry
1995-1998	B.S. Biochemistry, University of Washington Advisor: Craig Beeson

ACADEMIC EMPLOYMENT AND POSITIONS

2022-Present	Co-Director, UGA Cancer Center
2022-Present	Interim Department Head, University of Georgia Department of Pharmaceutical and Biomedical Sciences
2022-Present	University of Georgia, Full Prof. with Tenure Department of Pharmaceutical and Biomedical Sciences College of Pharmacy
2021-2022	Interim Associate Department Head, University of Georgia Department of Pharmaceutical and Biomedical Sciences
2020-2021	Interim Assistant Department Head, University of Georgia

	Department of Pharmaceutical and Biomedical Sciences
2019-Present	Dr. Samuel C. Benedict Professor, University of Georgia College of Pharmacy
08/2017-2022	University of Georgia, Associate Prof. with Tenure Department of Pharmaceutical and Biomedical Sciences College of Pharmacy
01/2017-Present	Adjunct Associate Professor, University of Georgia Departments of Chemistry and Biochemistry and Molecular Biology
08/2010-2016	Assistant Professor (tenure track), University of Georgia Department of Pharmaceutical and Biomedical Sciences
08/2010-2016	Adjunct Assistant Professor, University of Georgia Department of Biochemistry and Molecular Biology
06/2009-06/2010	Harvard University, Research Associate Department of Chemistry and Chemical Biology
01/2006-05/2009	Harvard University, American Cancer Society Postdoctoral Fellow Department of Chemistry and Chemical Biology
09/2000-12/2005	Univ. of CA San Diego, American Heart Association Predoctoral Fellow Department of Chemistry and Biochemistry
08/1998-08/2000	Puget Sound Blood Bank, Seattle, WA; Research Technologist II Division of Research, Genomic Research laboratory
07/1997-08/1998	University of Washington, Undergraduate Researcher Department of chemistry

AWARDS AND HONORS

1. *Session Chair*, 27th American Peptide Symposium, Whistler, Canada, June 2022
2. *Women's Leadership Fellow*, Selected by UGA Provost, 2021-2022
3. *Session Chair*, GRC on Chemistry and Biology of Peptides, Ventura, CA, March 2020
4. Endowed Professorship (Dr. Samuel C. Benedict Professor), 2019-Present
5. *Elected Member of Council*, American Peptide Society, 2019-2025
6. *Session Chair*, 26th American Peptide Symposium, Monterey, CA, June 2019
7. *Guest Editor,* Special Symposium-in-Print issue "Constrained Peptides and Biological Targets" for *Bioorganic and Medicinal Chemistry,* March 2018
8. *Session Chair, Gordon Research Conference on Phosphorylation and G-Protein*

Mediated Signaling Networks, Biddeford, ME, June 2016

9. *Session Chair*, 24th American Peptide Symposium, Orlando, FL, June 2015
10. *Elected Member of the Nominating Committee*, American Peptide Society, 2015-2017
11. *Keynote Speaker*, 12th Annual Biochemistry Retreat, University of Kassel, Wesendorf, Germany, January 22, 2015
12. *Session Chair*, 23rd American Peptide Symposium/ 6th Annual International Peptide Symposium, Waikoloa, HI, June 2013
13. *Faculty Summer Research Support for Outstanding Faculty*, Office of the Provost, University of Georgia, 2013
14. *Nominated Member,* Career Development and Mentoring Committee, ASPET, 2013-2016
15. *Session Chair*, Gordon Research Conference on Bioorganic Chemistry, Andover, NH, June 2011
16. *FASEB MARC Travel Awardee,* Sponsored by FASEB, EB 2011, Washington D.C., April 2011
17. *NCI Transition Career Development Award,* NCI K22 Award, 2011-2014
18. *Scholar-In-Training Award*, AACR-ACS Chemistry in Cancer Research, New Orleans, LA, 2009
19. *Scholar-In-Training Award*, AACR-ACS Chemistry in Cancer Research, San Diego, CA, 2007
20. *Carl. F. Storm Underrepresented Minority Fellowship*, Gordon Research Conference on Molecular Pharmacology, Lucca, Italy, 2005
21. *Postdoctoral Fellowship*, American Cancer Society, 2006-2009
22. *Travel Award*, Third International Conference on Structural Biology, Singapore, 2004
23. *Predoctoral Fellowship*, American Heart Association, 2003-2005
24. *Best Poster Award*, Molecular and Theoretical Biophysics Symposium, San Diego, CA, 2003
25. *Predoctoral Fellowship*, NSF MASEM Program, 2000-2001
26. *Outstanding Teaching Assistant Award*, Organic Chemistry Laboratory Teach Assistant, UCSD, 2000

U.S. AND INTERNATIONAL PATENTS

1. "Ligation of Stapled Polypeptides." Verdine GL and **Kennedy EJ.** (2015) 9,458,189 issued 10/4/2016
2. "EGFR targeting compounds and methods of use thereof." **Kennedy EJ** and Hanold LE. (2015) PCT/US2015/35621
3. "Chemically-stabilized A Kinase Anchoring Protein (AKAP) peptide disruptors." **Kennedy EJ,** Wang Y and Taylor SS. (2015), PCT/US2014/047236 and WO 2015/010048 A1
4. "Stably linked peptides and methods for their use." **Kennedy EJ** and Hanold LE. (2014) USSN 61/774,928

5. “Antiparasitic chemically-stabilized peptide disruptors.” **Kennedy EJ** and Peterson DS. (2014) USSN 61/866,580
6. “Disruption of the WAVE3 (WASF3) protein complex.” **Kennedy EJ.** (2015) USSN 62/144,631
7. “Disruption of the WAVE3 protein complex for suppression of invasion and metastasis.” **Kennedy EJ.** (2017) 15/565,229
8. “Chemically-stabilized allosteric modulators of Leucine-Rich Repeat Kinase 2 (LRRK2)” **Kennedy EJ,** Kortholt A and Helton LG. (2021) WO 2021/173802 A1

PUBLICATIONS

Publications, 2010-Present (* indicates corresponding author)

1. Bendzunas GN, Limaye AJ, Whittaker M, Cowell J, **Kennedy EJ**.* (2022) “Targeting the WASF3 complex to suppress metastasis.” *Pharmacological Research,* 182:106302, DOI: 10/1016/j.phrs.2022.106302 (PMID: 35691539).

2. Byrne DP, Omar MH, **Kennedy EJ**, Eyers PA, Scott JD. (2022) “Biochemical analysis of AKAP-anchored PKA signaling complexes.” Methods Mol Biol, 2483:297-317, DOI: 10.1007/978-1-0716-2245-2_19 (PMID: 35286684).

3. Singh R, Soliman A, Guaitoli G, Stormer E, von Zweydorf F, Maso TD, Van Rillaer L, Schmidt S, Chatterjee D, Pardon E, Knapp S, **Kennedy EJ**, Steyaert J, Herberg FW, Kortholt A, Gloeckner CJ, Versees W. (2022) “Nanobodies as allosteric modulators of Parkinson’s disease-associated LRRK2.” *Proc. Nat. Acad. Sci.,* 119: e2112712119, DOI: 10.1073/pnas.2112712119 (PMID: 35217606).

4. Limaye AJ, Bendzunas GN, Whittaker M, LeClair TJ, Helton LG, **Kennedy EJ.*** (2022) “In silico optimized stapled peptides targeting WASF3 in breast cancer,” *ACS Med Chem Letters,* DOI: 10.1021/acsmedchemlett.1c00627 (PMID: 35450347).

5. Helton LG, Rideout HJ, Herberg FW, **Kennedy EJ.*** (2022) “Leucine Rich Repeat Kinase 2 (LRRK2) Peptide Modulators: Recent Advances and Future Directions.” *Peptide Science,* 114:e24251, DOI: 10.1002/pep2.24251
   **Featured on Cover

6. Helton LG, Soliman A, von Zweydorf F, Kentros M, Manschwetus JT, Hall S, Gilsbach BK, Ho F, Athanasopoulos P, Singh R, LeClair TJ, Versees W, Raimondi F, Herberg FW, Gloeckner CJ, Rideout HJ, Kortholt A*, **Kennedy EJ**.* (2021) “Allosteric inhibition of Parkinson’s-linked LRRK2 by constrained peptides.” *ACS Chem Biol,* 16:2326-2338, DOI: 10.1021/acschembio.1c00487 (PMID: 34496561).

7. Limaye AJ, Bendzunas GN, **Kennedy EJ.*** (2021) “Targeted disruption of PKC from AKAP signaling complexes.” *RSC Chem Biol,* 2:1227-1231, DOI: 10.1039/d1cb00106j (PMID: 34458835).

8. Lasonder E, More K, Singh S, Haidar M, Bertinetti D, **Kennedy EJ**, Herberg FW, Holder T, Langsley G, Chitnis C. (2021) “cAMP-dependent signalling pathways as potential targets for inhibition of *Plasmodium falciparum* blood stages.” *Frontiers in Microbiology,* 12:684005, DOI: 10.3389/fmicb.2021.684005 (PMID: 34108954).

9. Baffi TR, Lorden G, Wozniak JM, Feichtner A, Yeung W, Kornev AP, King CC, Del Rio JC, Limaye AJ, Bogomolovas J, Gould CM, Chen J, **Kennedy EJ**, Kannan N, Gonzalez DJ, Stefan E, Taylor SS, Newton, AC. (2021) "mTORC2 controls the activity of PKC and Akt by phosphorylating a conserved TOR-interaction motif." *Science Signaling,* 14: 678, DOI: 10.1126/scisignal.abe4509 (PMID: 33850054).

10. Peng H, Cassel J, McCracken DS, Prokop JW, Sementino E, Cheung M, Collop PR, Polo A, Joshi S, Mandell JP, Ayyanathan K, Hinds D, Malkowicz SB, Harbour JW, Bowcock AM, Salvino J, **Kennedy EJ**, Testa JR, Rauscher FJ III. (2021) "Kinetic characterization of ASXL1/2-mediated allosteric regulation of the BAP1 deubiquitinase." *Mol. Cancer Research*, DOI: 10.1158/1541-7786.MCR-20-0080 (PMID: 33731362).

11. Helton L.G., Limaye A.J., Bendzunas G.N., **Kennedy E.J.*** (2020). "Novel Stabilized Peptide Inhibitors of Protein Kinases." In: Shapiro P. (eds) Next Generation Kinase Inhibitors. Springer, Cham. DOI: 10.1007/978-3-030-48283-1_7.

12. Ding K, McGee-Lawrence ME, Kaiser H, Sharma AK, Pierce JL, Irsik DL, Bollag WB, Xu J, Zong Q, Hill W, Shi X-M, Fulzele S, **Kennedy EJ**, Elsalanty M, Hamrick MW and Isales CM (2020). "Picolinic acid, a tryptophan oxidation product, does not impact bone mineral density but increases marrow adiposity." *Experimental Gerontology,* 133:110885, DOI: 10.1016/j.exger.2020.110885 (PMID: 32088397).

13. Helton LG and **Kennedy EJ*** (2020). "Targeting plasmodium with constrained peptides and peptidomimetics." *IUBMB Life,* DOI: 10.1002/iub.2244 (PMID: 32037730).

14. Manschwetus JT, Bendzunas GN, Limaye AJ, Knape MJ, Herberg FW*, **Kennedy EJ*** (2019) "A stapled peptide mimic of the pseudosubstrate inhibitor PKI inhibits Protein Kinase A." *Molecules,* 24(8):1567, DOI: 10.3390/molecules24081567 (PMID: 31009996).

15. Cao Z, Singh B, Li C, Markham NO, Carrington LJ, Franklin JL, Graves-Deal R, **Kennedy EJ**, Goldenring JR and Coffey EJ (2019) "PKA-mediated phosphorylation of NKD2 stimulates cell-surface delivery of TGF-a for EGFR transactivation." *Traffic*, 20(5): 357-368, DOI: 10.1111/tra.12642 (PMID: 30941853).

16. Flaherty B, Ho TG, Schmidt S, Herberg FW, Peterson D, **Kennedy EJ*** (2019) "Targeted Inhibition of Plasmodium falciparum Calcium-Dependent Protein Kinase 1 With A Constrained J Domain-Derived Disruptor Peptide." *ACS Infect Dis.*, 5(4): 506-514, DOI: 10.1021/acsinfecdis.8b00347. (PMID: 30746930).

17. **Kennedy EJ*** (2018) "Constrained peptides and biological targets." *Bioorg. Med. Chem*., 26: 1117. DOI: 10.1016/j.bmc.2018.02.046 (PMID: 29519596).

18. Burgess SG, Mukherjee M, Sabir S, Joseph N, Gutiérrez-Caballero C, Richards MW, Huguenin-Dezot N, Chin JW, **Kennedy EJ**, Pfuhl M, Royle SJ, Gergely F, Bayliss R. (2018) "Mitotic spindle association of TACC3 requires Aurora-A-dependent stabilization of a cryptic a-helix." *EMBO J*., 37(8). pii: e97902. DOI: 10.15252/embj.201797902 (PMID: 29510984).

19. Bendzunas NG, Dörfler S, Autenrieth K, Bertinetti D, Machal EMF, **Kennedy EJ***, Herberg FW*. (2018) "Investigating PKA-RII specificity using analogs of the PKA:AKAP peptide inhibitor STAD-2." *Bioorg. Med. Chem.,* 26(6):1174-1178. DOI: 10.1016/j.bmc.2018.02.001 (PMID: 29449124).

20. Haidar M, de Laté PL, **Kennedy EJ**, Langsley G. (2018) “Cell penetrating peptides to dissect host-pathogen protein-protein interactions in Theileria-transformed leukocytes.” *Bioorg. Med. Chem.,* 26(6):1127-1134. DOI: 10.1016/j.bmc.2017.08.056 (PMID: 28917447).

21. Fulton MD, Hanold LE, Ruan Z, Patel S, Beedle AM, Kannan N, **Kennedy EJ***. (2018) “Conformationally constrained peptides target the allosteric kinase dimer interface and inhibit EGFR activation.” *Bioorg. Med. Chem.,* 26(6):1167-1173. DOI: 10.1016/j.bmc.2017.08.051 (PMID: 28911855).

22. Egan JB, . . . , **Kennedy EJ**, Klee EW, Borad MJ, Fernandez-Zapico ME. (2017) “Functional analysis of exome sequencing-derived variants identifies a novel constitutively active FGFR2 mutant in cholangiocarcinoma.” *Journal of Clinical Oncology: Precision Oncology,* DOI: 10.1200/PO.17.00018.

23. El Refaey M, Lawrence-McGee ME, Fulzele S, **Kennedy EJ**, Bollag WB, Elsalanty M, Zhong Q, Ding KH, Bendzunas NG, Shi XM, Xu J, Hill WD, Johnson MH, Hunter M, Pierce JL, Yu K, Hamrick MW, Isales CM. (2017) “Kynurenine, a tryptophan metabolite that accumulates with age, induces bone loss.” *Journal of Bone and Mineral Research,* DOI: 10.1002/jbmr.3224 (PMID 28727234).

24. Cowell JK, Teng Y, Bendzunas NG, Ara R, Arbab AS, **Kennedy EJ***. (2017) “Suppression of breast cancer metastasis using stapled peptides targeting the WASF regulatory complex.” *Cancer Growth Metastasis,* DOI 10.1177/1179064417713197 (PMID: 28680267).

25. Mo GC, Ross B, Hertel F, Manna P, Yang X, Greenwald E, Booth C, Plummer AM, Tenner B, Chen Z, Wang Y, **Kennedy EJ**, Cole PA, Fleming KG, Palmer A, Jimenez R, Xiao J, Dedecker P, Zhang J. (2017) “Genetically-encoded biosensors for visualizing live-cell biochemical activity at superresolution.” *Nature Methods,* 14(4): 427-434, DOI: 10.1038/nmeth.4221 (PMID: 28288122).

26. Hanold LE, Fulton MD, **Kennedy EJ*.** (2017) “Targeting kinase signaling pathways with constrained peptide scaffolds.” *Pharmacology and Therapeutics,* 173: 159-170, DOI: 10.1016/j.pharmthera.2017.02.014 (PMID: 28185915).

27. Haidar M, Lombes A, Bouillaud F, **Kennedy EJ** and Langsley G (2017). “HK2 recruitment to phospho-BAD prevents its degradation promoting Warburg glycolysis by Theileria-transformed leukocytes.” *ACS Infectious Diseases,* 3(3):216-224, DOI: 10.1021/acsinfecdis.6b00180 (PMID: 28086019).

28. Haidar M, Ramdani G, **Kennedy EJ** and Langsley G. (2017) “PKA and apicomplexan parasite diseases.” *Horm. Metab. Res.,* 48: 1-5, DOI 110.1055/s-0042-118459 (PMID: 27835919).

29. Teng Y*, Qin H, Bahassan A, Bendzunas NG, **Kennedy EJ***, Cowell JK*. (2016) “The WASF3-NCKAP1-CYFIP1 Complex Is Essential for Breast Cancer Metastasis.” *Cancer Res*earch, 76(17):5133-42, DOI: 10.1158/0008-5472.CAN-16-0562 (PMID: 27432794).

30. Gotz F, Roske Y, Schulz MS, Autenrieth K, Bertinetti D, Faelber K, Zuhlke K, Kreuchwig A, **Kennedy EJ**, Krause G, Daumke O, Herberg FW, Heinemann U and Klussmann E.

(2016) “AKAP18:PKA-RIIa structure reveals crucial anchor points for recognition of regulatory subunits of PKA.” *Biochemical J.,* 473(13):1881-94, DOI 10.1042/BCJ20160242 (PMID: 27102985).

31. Mohanty S, Oruganty K, Kwon A, Byrne DP, Ferries S, Ruan Z, Hanold LE, Katiyar S, **Kennedy EJ,** Eyers PA and Kannan N. (2016) “Hydrophobic core variations provide a structural framework for tyrosine kinase evolution and functional specialization.” *PLoS Genetics*, 12: e1005885, DOI 10.1371/journal.pgen.1005885 (PMID: 26925779).

32. Bavencoffe A, Li Y, Wu Z, Yang Q, Herrera J, **Kennedy EJ**, Walters ET, Dessauer CW. (2016) “Persistent Electrical Activity in Primary Nociceptors after Spinal Cord Injury Is Maintained by Scaffolded Adenylyl Cyclase and Protein Kinase A and Is Associated with Altered Adenylyl Cyclase Regulation.” *J. Neuroscience*, 36(5):1660-8, DOI 10.1523/JNEUROSCI.0895-15.2016 (PMID: 26843647).

33. Teng Y, Bahassan A, Dong D, Hanold LE, Ren X, **Kennedy EJ**, Cowell JK. (2016) “Targeting the WASF3-CYFIP1 Complex Using Stapled Peptides Suppresses Cancer Cell Invasion.” *Cancer Research*, 76(4):965-73, DOI 10.1158/0008-5472.CAN-15-1680 (PMID: 26676744).

34. Autenrieth K, Bendzunas G, Bertinetti D, Herberg FW*, **Kennedy EJ***. (2016) “Defining A-Kinase Anchoring Protein (AKAP) Specificity for Protein Kinase A Subunit RI (PKA-RI).” *ChemBioChem*, 17(8):693-7. DOI 10.1002/cbic.201500632 (PMID: 26611881).

35. Nguyen HT, Jia W, Beedle AM, **Kennedy EJ**, Murph MM. (2015) “Lysophosphatidic Acid Mediates Activating Transcription Factor 3 Expression Which Is a Target for Post-Transcriptional Silencing by miR-30c-2-3p.” *PLoS One,* 10(9):e0139489, DOI 10.1371/journal.pone.0139489 (PMID: 26418018).

36. **Kennedy EJ*** and Kannan N. (2015) “Dialing in EGFR signaling.” *Chemistry and Biology,* 22(6): 687-8, DOI 10.1016/j.chembiol.2015.06.001 (PMID: 26091165).

37. Flaherty BR, Wang Y, Trope EC, Ho TG, Muralidharan V, **Kennedy EJ*** and Peterson DS*. (2015) “The Stapled AKAP disruptor peptide STAD-2 displays antimalarial activity through a PKA-independent mechanism.” *PLoS One,* 10(5): e0129239, DOI 10.1371/journal.pone.0129239 (PMID: 26010880).

38. **Kennedy EJ** and Scott JD. (2015) “Selective disruption of the AKAP signaling complexes.” *Methods in Molecular Biology*, 1294: 137-50, DOI 10.1007/978-1-4939-2537-7_11 (PMID: 25783883).

39. Hermann JS, Skroblin P, Bertinetti D, Hanold LE, von der Heide EK, Wagener E-M, Zenn H-M, Klussmann E, **Kennedy EJ** and Herberg FW. (2015) “Neurochondrin is an atypical RIIalpha-specific A-kinase anchoring protein.” *Biochim. Biophys. Acta,* 1854:1667-75, DOI 10.1016/j.bbapap.2015.04.018 (PMID: 25916936).

40. Mohanty S, **Kennedy EJ**, Herberg FW, Hui R, Taylor SS, Langsley G and Kannan N. (2015) “Structural and evolutionary divergence of cyclic nucleotide binding domains in eukaryotic pathogens: Implications for drug design.” *Biochim. Biophys. Acta,* 1854:1575-85. DOI 10.1016/j.bbapap.2015.03.012. (PMID: 25847873).

41. Hanold LE, Watkins, CP, Liaw P, Beedle AM and **Kennedy EJ***. (2015) "Design of a selenylsulfide-bridged EGFR dimerization arm mimic." *Bioorg. Med. Chem.,* 23(12):2761-6. DOI 10.1016/j.bmc.2015.03.040 (PMID: 25840798).

42. Hanold LE, Oruganty K, Ton NT, Beedle AM, Kannan N and **Kennedy EJ***. (2015) "Inhibiting EGFR dimerization using triazolyl-bridged dimerization arm mimics." *PLoS One*, 10(3): e0118796, DOI 10.1371/journal.pone.0118796 (PMID: 25790232).

43. Wang Y, Ho TG, Franz E, Hermann JS, Smith FD, Hehnly H, Esseltine JL, Hanold LE, Murph MM, Bertinetti D, Scott JD, Herberg FW and **Kennedy EJ***. (2015) "PKA-type I Selective constrained peptide disruptors of AKAP complexes." *ACS Chemical Biology*, 10(6):1502-10. DOI 10.1021/acschembio.5b00009 (PMID 25765284).

44. El Refaey M, Watkins CP, **Kennedy EJ**, Chang A, Zong Q, Ding K-H, Shi X-M, Xu J, Bollag WB, Hill WD, Johnson M, Hunter M, Hamrick MW and Isales CM. (2015) "Oxidation of the aromatic amino acids tryptophan and tyrosine disrupts their anabolic effects on bone marrow-derived mesenchymal stem cells." *Mol. Cell. Endrocinol.,* 410:87-96. DOI 10.1016/j.mce.2015.01.034 (PMID: 25637715).

45. **Kennedy EJ***. (2014) "EMBO conference series: Chemical Biology 2014." *ChemBioChem*. 15(18):2783-7. DOI 10.1002/cbic.201402527. (PMID: 25318996).

46. Fortunato MJ, Ball CE, Hollinger K, Patel NB, Modi JN, Rajasekaran V, Nonneman DJ, Ross JW, **Kennedy EJ,** Selsby JT, Beedle AM. (2014) "Development of rabbit monoclonal antibodies for detection of alpha-dystroglycan in normal and dystrophic tissue." *PLoS One,* 9(5):e97567. doi: 10.1371/journal.pone.0097567 (PMID: 24824861).

47. **Kennedy EJ***. (2014) "Biological Drug Products: Development and Strategies." Edited by Wei Wang and Manmohan Singh. *ChemMedChem.* 9(12): 2814-2815.

48. Wang Y, Ho TG, Bertinetti D, Neddermann M, Franz E, Mo GC, Schendowich LP, Sukhu A, Spelts RC, Zhang J, Herberg FW, **Kennedy EJ**. * (2014) "Isoform-selective disruption of AKAP-localized PKA using hydrocarbon stapled peptides." *ACS Chemical Biology,* 9(3):635-42. doi: 10.1021/cb400900r (PMID: 24422448).

Publications, Prior to 2010

49. Yang J, **Kennedy EJ**, Wu J, Deal MS, Pennypacker J, Ghosh G, and Taylor SS. (2009) "Contribution of non-catalytic core residues to activity and regulation in PKA." *J. Biol. Chem.,* 284: 6241-6248, (PMCID 19122195).

50. **Kennedy EJ**, Yang J, Pillus L, Taylor SS, and Ghosh G. (2009) "Identifying critical non-catalytic residues that modulate PKA activity." *PLoS One,* 4:e4746, (PMCID 19270744).

51. **Kennedy EJ**, Ghosh G, and Pillus L. (2008) "Identification of functionally distinct regions that mediate biological activity of the Protein Kinase A homolog Tpk2." *J. Biol. Chem.,* 283: 1084-1093, (PMCID 17971450).

52. **Kennedy EJ**, Pillus L, and Ghosh G. (2005) "Pho5p and newly identified E-NPPs regulate nucleotide phosphate metabolism in *S. cerevisiae*," *Eukaryotic Cell,* 11: 1892-1901, (PMCID 16278456).

53. Gaur LK, Nitta Y, **Kennedy E**, Lernmark A, Nelson K, Allen M, Nepom G. (2002) "Induction of islet allotolerance in nonhuman primates." *Ann. NY Acad. Sci.,* 958, 199-203, (PMCID 12021106).

54. Gaur LK, **Kennedy E,** Nitta Y, Nepom G, Nelson K, Allen M, Lernmark A. (2002) "Induction of donor-specific tolerance to islet allografts in nonhuman primates." *Ann. NY Acad. Sci.,* 958, 194-198, (PMCID 12021105).

INVITED PRESENTATIONS

CONFERENCE LECTURES, 2010-Present

1. Confirmed Speaker: Canadian Chemistry Conference and Exhibition, Vancouver, Canada, June 4-8, 2023
2. Confirmed Speaker: GRC Chemistry and Biology of Peptides, Oxnard, CA, Oct 30-Nov 4, 2022
3. Foldamers NYU Meeting, New York, NY, July 26-29, 2022, "Doubly-constrained peptides targeting LRRK2"
4. UGA-Emory Joint Cancer Center Retreat, Stone Mountain, GA, July 17-18, 2022, "Targeting the WASF3 regulatory complex with constrained peptides"
5. LRRK2 Central, July 7, 2022, "Targeting LRRK2 with constrained peptides"
6. Biochemical Society LRRK2 Meeting, Lille, France, June 28-July 1, 2022, "Targeting LRRK2 with constrained peptides"
7. Pacifichem 2021, December 15-20, 2021, ""Allosteric Targeting of LRRK2"
8. Experimental Biology 2021, April 27-30, 2021, "Targeting LRRK2 in Parkinson's"
9. 2021 American Chemical Society National Meeting, April 5-30, 2021 "Design of constrained peptides targeting the tumor metastasis regulator WASF3"
10. Michael J. Fox Foundation LRRK2 Consortium, March 24, 2021 "Allosteric targeting of the Parkinson's related kinase LRRK2"
11. American Peptide Society eSeminar Series, Feb. 16, 2021 "Allosteric Targeting of Kinases using constrained peptides"
12. 17th Annual Biochemistry Retreat, Universitat Kassel, Wesendorf, Germany, Jan. 8-11, 2020
13. Moving Breast Cancer Treatments Forward Symposium, Jayne Koskinas Ted Giovanis Foundation for Health and Policy (JKTG Foundation), Washington, DC, Oct. 30, 2019
14. 10th Inhibitors of Protein Kinases Conference, Warsaw, Poland, Sept. 14-18, 2019 "Targeted inhibition of the plasmodium kinase CDPK1 using constrained peptides"
15. 26th American Peptide Symposium, Monterey, CA, June 22-27, 2019 "Targeting LRRK2 Dimerization in Parkinson's"
16. 2019 American Chemical Society National Meeting, Orlando, FL, Mar. 31-April 4, 2019 "Allosteric kinase inhibition via disrupted dimerization"
17. 16th Annual Biochemistry Retreat, Universitat Kassel, Wesendorf, Germany, Jan. 9-12, 2019, "Targeted degradation of the DnaJ1-PRKACA chimera"

18. ASBMB Kinases and Pseudokinases, San Diego, CA, Dec 9-12, 2018, “Targeted inhibition of LRRK2 using constrained peptides”
19. Greater Atlanta Chemical Biology Symposium, Atlanta, GA, April 21, 2018, "Targeting kinase regulation with constrained peptide Scaffolds”
20. 15th Annual Biochemistry Retreat, Universitat Kassel, Wesendorf, Germany, Jan. 10-13, 2018, “Targeting WASF3 in Tumor Metastasis”
21. Pharmacology 2017 and the British Pharmacological Society, London, England, Dec. 11-13, 2017, "Targeting kinase regulation with constrained peptides”
22. LRRK2 Workshop, Kassel, Germany, Sept. 14-15, 2017, “Targeted disruption of kinase dimerization for allosteric inhibition”
23. 9th Inhibitors of Protein Kinases Conference, Warsaw, Poland, September 17-21, 2017 “Targeting AKAP signaling complexes with constrained peptide scaffolds”
24. 25th American Peptide Symposium, Whistler, Canada, June 17-22, 2017 “Targeting AKAP signaling complexes with constrained peptide scaffolds”
25. 14th Annual Biochemistry Retreat, Universitat Kassel, Wesendorf, Germany, Jan. 11-14, 2017, “Pharmacological inhibition of kinase activation”
26. 5th International Meeting on Anchored cAMP Signaling, Zermatt, Switzerland, Oct. 6-9, 2016, “Targeting AKAP complexes using constrained peptide scaffolds”
27. Gordon Research Conference on Phosphorylation and G-Protein Mediated Signaling Networks, Biddeford, ME, June 5-10, 2016, “Targeting allosteric activation of EGFR”
28. Experimental Biology 2016, San Diego, CA, April 2-6, 2016, “Developing novel chemical biology strategies to synthetically disrupt protein-protein interactions”
29. 13th Annual Biochemistry Retreat, Universitat Kassel, Wesendorf, Germany, Jan. 20-23, 2016, “Targeting regulation and allosteric activation of protein kinases”
30. ASBMB Special Symposia on Kinases and Pseudokinases, San Diego, CA, Dec. 5-8, 2015, “Targeting allosteric activation of EGFR”
31. 4th International Workshop on cAMP signaling, PKA and phosphodiesterases, Erciyes University, Kayseri, Turkey, September 3-5, 2015, “Dissecting AKAP signaling complexes”
32. UGA Cancer Center Retreat, Athens, GA, October 30, 2015, “Chemical biology approaches to study kinase allostery”
33. Drug Discovery and Therapy World Congress, Boston, MA, July 21-25, 2015, “Inhibiting EGFR dimerization using triazolyl-bridged dimerization arm mimics”
34. Research Symposium Honoring Gourisankar Ghosh, UCSD, San Diego, CA, June 18-20, 2015, “Chemical biology approaches to study kinase allostery”
35. 12th Annual Biochemistry Retreat, Universitat Kassel, Wesendorf, Germany, Jan. 21-24, 2015, "Design of chemically modified peptides to study regulation of protein kinases”
36. EMBO Chemical Biology Meeting, EMBL Heidelberg, Heidelberg, Germany, Aug. 20-23, 2014, "Isoform-selective disruption of PKA-AKAP complexes and its effects as an antimalarial"
37. 8th International Conference on Inhibitors of Protein Kinases, Warsaw, Poland, Sept. 21-25, 2014, "Stabilized peptides targeting AKAP anchoring complexes"

38. Cyclic Nucleotide Signaling Workshop, CNRS, Paris, France, Dec. 15-17, 2013, “Design of conformationally constrained AKAP inhibitor peptides”
39. 4th International Meeting on Anchored cAMP Signaling Pathways, Denver, CO, October 4-6, 2013, “Design of conformationally constrained AKAP inhibitor peptides”
40. UGA-GRU Joint Cancer Center Retreat, Lake Oconee, GA, Nov. 1, 2013, “Designing peptide-based allosteric kinase effectors”
41. Experimental Biology 2012, San Diego, CA, April 21-25, 2012, “Targeting EGFR in cancer: cancer drug discovery”
42. Southern Translational Education and Research Conference, Augusta, GA, Sept. 16, 2011, “Design of novel peptide-based inhibitors targeting EGFR”

ACADEMIC SEMINARS, 2010-Present

1. University of Iowa, Dept. of Pharmaceutical Sciences and Experimental Therapeutics, March 23, 2022, “Targeting PPIs with Constrained Peptides”
2. University of Leeds, Leeds, England, Asbury Lecture, Feb. 28, 2020, “Allosteric inhibition of kinases with constrained peptides”
3. University of Groningen, Groningen, Netherlands, Dept. of Cell Biochemistry, Feb. 26, 2020, “Allosteric inhibition of kinases with constrained peptides”
4. University of South Carolina, Dept. of Drug Discovery and Biomedical Sciences, Oct. 8, 2019, “Allosteric kinase inhibition with constrained peptides”
5. Oregon Health Sciences University, Dept. of Pharmacology, Oct. 18, 2018, “Targeting kinases with constrained peptide scaffolds”
6. University of Georgia, Dept. of Chemistry, Oct. 4, 2018, “Targeting kinases with constrained peptide scaffolds”
7. University of Kassel, Kassel, Germany, December 12, 2017, “Targeting kinase regulation with constrained peptide scaffolds”
8. University of Texas Medical Center, Houston, Texas, November 21, 2016, “Targeting Kinase Regulation Using Chemical Biology”
9. Max Delbruck Centrum, Berlin, Germany, October 9-10, 2016, “Stapled peptides for pharmacological interference with cAMP signaling"
10. The Wistar Institute, Program in Gene Expression and Regulation, Philadelphia, PA, March 9, 2016, “Targeting kinase regulation using constrained peptide scaffolds”
11. University of Iowa, Div. of Medicinal and Natural Products Chemistry and Molecular and Cellular Biology, Iowa City, IA, March 3, 2016, “Targeting kinase regulation using constrained peptide scaffolds”
12. Max Planck Institute for Molecular Physiology, Dortmund, Germany, Jan. 19, 2015, “Chemical Biology Tools to Study Kinase Regulation"
13. San Diego State University, Dept. of Chemistry, Dec. 12, 2014, "Design of chemically modified peptides to study regulation of protein kinases”
14. University of California, San Diego, Dept. of Chemistry and Biochemistry, Dec. 11, 2014, "Design of chemically modified peptides to study regulation of protein kinases”

15. University of Vermont, Dept. of Pharmacology, Aug. 28, 2014, "Design of chemically modified peptides to study regulation of protein kinases"
16. Mayo Clinic, Dept. of Oncology, Rochester, MN, March 20, 2014, "Design of chemically modified peptides to study kinase regulation"
17. *European Molecular Biology Laboratory (EMBL),* Dept. of Cell Biology and Biophysics, *Heidelberg, Germany, December 18, 2013,* "Designing modified peptides to study kinase cell signaling"
18. University of Georgia, Dept. of Infectious Diseases, Sept. 16, 2013, "Designing modified peptides to study kinase cell signaling"
19. University of Virginia, School of Medicine, Dept. of Molecular Physiology and Biophysics, Charlottesville, VA, March 11, 2013, "Designing modified peptides to study kinase cell signaling"
20. Georgia Regents University, Inst. of Molecular Medicine and Genetics, Augusta, GA, Feb. 25, 2013, "Designing modified peptides to study kinase cell signaling"
21. University of Georgia, Office of the Vice President for Research, April 8, 2013, "Writing a winning NIH K award application"
22. University of Georgia, Center for Drug Discovery, March 5, 2013, "Designing modified peptides to study kinase cell signaling"
23. University of Georgia, Dept. of Physiology and Pharmacology, March 26, 2012, "Spatiotemporal regulation of kinase signaling in cancer"
24. *University of Georgia, Biochemistry Graduate Student Association Seminar Series, Jan. 4, 2011, "Synthetic biologics: new approaches to temporally alter cell signaling"*
25. Foundation for Applied Molecular Evolution, Gainesville, FL, July 15, 2011, "Synthetic Biologics: New Approaches to Target Cell Signaling"
26. *University of Georgia, Dept. of Biochemistry and Molecular Biology, Sept. 8, 2010, "Chemical Stabilization of alpha helices, beta sheets, and semi-synthetic proteins"*

SUMMARY OF TEACHING RESPONSIBILITIES, 2016-Present

Semester	Course	Total Enrollment	Percent Responsible	Credit Hours
Fall 2021	**PHRM4060 – Medicinal Chemistry II**	143	87	2
Fall 2021	PHRM9000 – Doctoral Research	4	100	12
Fall 2021	BCMB4960R – Undergraduate Research	1	100	4
Spring 2021	**PHRM8020 – Mol. Pharm. of Disease and Therapeutics**	6	30	4
Spring 2021	PHRM9000 – Doctoral Research	5	100	11.6
Spring 2021	BCMB4970R – Undergraduate Research	1	100	4
Spring 2021	BCMB4980R – Undergraduate Research	1	100	4
Spring 2021	PHRM9300 – Doctoral Dissertation	1	100	3
Fall 2020	**PHRM4060 – Medicinal Chemistry II**	144	83	2
Fall 2020	PHRM9000 – Doctoral Research	3	100	12.3
Fall 2020	BCMB4960R – Undergraduate Research	1	100	4
Fall 2020	BCMB4970R – Undergraduate Research	1	100	4
Summer 2020	PHRM9000 – Doctoral Research	3	100	17.3
Spring 2020	PHRM3070 – Medicinal Chemistry I	1	100	2
Spring 2020	**PHRM8020 – Mol. Pharm. of Disease and Ther.**	18	30	4
Spring 2020	PHRM8010 – Structural Bio and Med Chem	8	6	4
Spring 2020	PHRM9000 – Doctoral Research	3	100	12

Spring 2020	BCMB4960R – Undergraduate Research	1	100	4
Spring 2020	BCMB4970R – Undergraduate Research	1	100	4
Fall 2019	**PHRM4060 – Medicinal Chemistry II**	131	83	2
Fall 2019	PHRM9000 – Doctoral Research	3	100	13.7
Fall 2019	BCMB4980R – Undergraduate Research	1	100	4
Summer 2019	PHRM9000 – Doctoral Research	3	100	18
Summer 2019	BCMB4980H – Undergraduate Research	1	100	4
Spring 2019	PHRM3070 – Medicinal Chemistry I	138	20	2
Spring 2019	PHRM8080 – Grantsmanship	15	5	3
Spring 2019	**PHRM8020 – Mol. Pharm. of Disease and Ther.**	14	30	4
Spring 2019	PHRM8010 – Structural Bio and Med Chem	5	6	4
Spring 2019	PHRM9000 – Doctoral Research	2	100	13
Spring 2019	BIOL4970H – Undergraduate Research	1	100	4
Spring 2019	CBIO4970H – Directed Laboratory Research	1	100	4
Fall 2018	**PHRM4060- Medicinal Chemistry II**	133	60	2
Fall 2018	PHRM 9000 – Doctoral Research	3	100	9
Fall 2018	BIOL4960H – Undergraduate Research	1	100	4
Fall 2018	CBIO4980H – Directed Laboratory Research	1	100	4
Summer 2018	PHRM9000 – Doctoral Research	2	100	18
Summer 2018	CBIO4980H – Directed Laboratory Research	1	100	4
Spring 2018	PHRM3070 – Medicinal Chemistry I	137	20	2
Spring 2018	PHRM8020 – Mol. Pharm. of Disease and Ther.	19	28	4
Spring 2018	PHRM8010 – Structural Bio and Med Chem	17	6	4
Spring 2018	PHRM8080 – Grantsmanship	5	5	3
Spring 2018	PHRM5980 – PharmD Research	4	100	4
Spring 2018	PHRM9000 – Doctoral Research	2	100	9
Spring 2018	PHRM7000 – Master's Research	1	100	4
Fall 2017	**PHRM4060 – Medicinal Chemistry II**	142	50	2
Fall 2017	PHRM5980 – PharmD Research	2	100	3
Fall 2017	BCMB4970R – Undergraduate Research	1	100	4
Fall 2017	CBIO4970H – Directed Laboratory Research	1	100	4
Fall 2017	PHRM7000 – Master's Research	1	100	9
Summer 2017	CBIO4960H – Directed Laboratory Research	1	100	4
Spring 2017	PHRM3070 – Medicinal Chemistry I	145	20	2
Spring 2017	PHRM8020 – Mol. Pharm. of Disease and Ther.	15	5	4
Spring 2017	PHRM8080 – Grantsmanship	7	5	3
Spring 2017	PHRM8010 – Structural Bio and Med Chem	5	7	4
Spring 2017	BCMB4960L – Undergraduate Research	1	100	4
Spring 2017	BCMB4970L – Undergraduate Research	1	100	4
Spring 2017	PHRM7000 – Master's Research	1	100	7
Fall 2016	**PHRM4060 – Medicinal Chemistry II**	137	50	2
Fall 2016	BCMB4960L – Undergraduate Research	1	100	4
Fall 2016	PHRM5980 – PharmD Research	1	100	3
Fall 2016	PHRM9000 – Doctoral Research	1	100	13
Summer 2016	PHRM9000 – Doctoral Research	1	100	18

SUMMARY OF PharmD TEACHING EVALUATIONS, 2016-Present

Course	**PHRM 3070 (Med Chem I)**				
(1-5 Scale; 1 strongly disagree - 5 strongly agree) **Year**	**2016**	**2017**	**2018**	**2019**	**2020**
Instructor was organized & prepared:	4.82	4.65	4.78	4.79	4.90
Clarity in presenting materials:	4.66	4.40	4.90	4.74	4.87
Stimulated interest in the topic:	4.47	4.09	4.74	4.90	4.97

Assessments were consistent and clear:	4.64	4.29	4.71	4.71	4.84
Professional interaction with students:	4.92	4.65	4.91	4.91	4.89
Average	**4.70**	**4.42**	**4.81**	**4.81**	**4.89**

Course	**PHRM 4060 (Med Chem II)**				
(1-5 Scale; 1 strongly disagree - 5 strongly agree) **Year**	**2016**	**2017**	**2018**	**2019**	**2020**
Instructor was organized & prepared:	4.88	4.83	4.92	4.93	4.91
Clarity in presenting materials:	4.88	4.71	4.87	4.87	4.89
Stimulated interest in the topic:	4.77	4.33	4.97	4.97	4.96
Assessments were consistent and clear:	4.87	4.83	4.86	4.92	4.90
Professional interaction with students:	4.88	4.89	4.95	4.97	4.96
Average	**4.86**	**4.72**	**4.91**	**4.93**	**4.92**

SUPERVISION OF RESEARCH

Supervision of Graduate Student Research (My Role as Primary Research Advisor)

Years	Graduate Student	Current Position
2011-2016	Laura E. Hanold, Ph.D.	Postdoctoral Fellow, U Florida
2011-2014	Yuxiao Wang, M.S.	Director of Research, Parkway Clinical Labs
2012-2019	Melody Fulton, Ph.D.	(changed labs in 2017) Res. Scientist, U Idaho
2017-2021	George N. Bendzunas	Postdoctoral Fellow, UGA
2017-2022	Leah G. Helton, Ph.D.	Research Scientist, Athira Pharma
2017-2022	Ameya Limaye	Research Scientist, SyntheX
2020-Present	Matt Whittaker	Continuing Ph.D. Student
2020-Present	Krista Barbour	Continuing Ph.D. Student

Graduate Student Advisory Committee Membership

Years	Graduate Student	Department at UGA
2012-2015	Briana Flaherty, Ph.D.	Infectious Diseases
2011-2015	Ha Nguyen, Ph.D	Pharmaceutical and Biomedical Sciences
2011-2013	Ranjan Behera, Ph.D.	Cellular Biology
2011-2014	Linna Yan, M.S.	Pharmaceutical and Biomedical Sciences
2012-2014	MD Fazlur Rahman, Ph.D.	Pharmaceutical and Biomedical Sciences
2010-2011	DJ Bernsteel, Ph.D.	Biochemistry and Molecular Biology
2011-2016	Catherine Sullenberger, Ph.D.	Cellular Biology
2012-2017	Kaitlyn Ledwich, Ph.D.	Pharmaceutical and Biomedical Sciences
2013-2019	Hyunjin Kwon, Ph.D.	Biochemistry and Molecular Biology
2012-2017	Yiwen Zhang, Ph.D.	Cellular Biology
2012-2018	Emily Carpinone, Ph.D.	Microbiology
2012-2018	Mohammed Alqinyah, Ph.D.	Pharmaceutical and Biomedical Sciences
2012-2018	Zhen Han, Ph.D.	Pharmaceutical and Biomedical Sciences
2012-2018	Liza Ngo, Ph.D.	Pharmaceutical and Biomedical Sciences
2012-2018	Ruan Zheng, Ph.D.	Biochemistry and Molecular Biology
2012-2018	Wided Missaoui, Ph.D.	Pharmaceutical and Biomedical Sciences
2013-2020	Charnel Byrnes, Ph.D.	Pharmaceutical and Biomedical Sciences
2015-2020	Sukhneeraj Kaur, Ph.D.	Pharmaceutical and Biomedical Sciences
2016-Present	Menbere Wendimu	Pharmaceutical and Biomedical Sciences
2018-2020	Dylon Stephens, M.S.	Pharmaceutical and Biomedical Sciences
2017-Present	Wayland Yeung	Biochemistry and Molecular Biology
2016-Present	Yueze Yang	Pharmaceutical and Biomedical Sciences
2019-Present	Safal Shrestha	Biochemistry and Molecular Biology

2021-Present	Autumn Tobin	Pharmaceutical and Biomedical Sciences

Supervision of Undergraduate Researchers

Years	Undergrad. Student	Current Position
2010-2011	Fenil Patel	Chief Resident, Sioux Fall Family Med. Residency
2011-2012	Viral Patel	Clinical Pharmacist Lead, Humana
2013	Nita Jain	Freelance writer
2011-2013	Norman Ton	Pharmacist, US Army
2012-2014	Christopher Watkins	PhD Student, U of Chicago
2013-2014	Avinash Sukhu	Lab Technician, Georgia State University
2013-2014	Lewis Schendowich	Resident Physician, St. Mary's Medical Center
2014-2015	Peter Cieszewski	Chemist, HB Fuller
2014-2015	Peter Liaw	Resident, University of Virginia School of Medicine
2016-2017	Tonya Ly	Compounder, Averr Aglow Skincare
2017	Patrick Humphreys	Graduate Student, Imperial College London
2017-2019	Ismar Miniel	Graduate Student, U Texas Austin
2018-2020	Scotty Hall	Medical Student, Augusta University
2020-Present	Tyler Moore	Medical Student, Mercer University
2020-Present	Jonathan Schulz	Medic training, Boston, MA
2021-Present	Nick Tsavaris	Continuing BS student
2022-Present	Nick Tawadrous	Continuing BS student

Supervision of PharmD Researchers and High School Students

Years	PharmD Student	Current Position
2011	Laura Cotter	Senior Manager of Ed., Discovery Gateway Museum
2012-2014	Tienhuei Grace Ho	Pharmacist, CHOA Scottish Rite
2012-2013	Raybun Spelts	Pharmacist, Hospital Authority of Miller County
2014	Huong Pham	Pharmacist, Emory University
2015-2016	Courtlyn Smith	Pharmacist Resident, Navicent Health
2015-2016	Michelle Vu	Pharmacist, Northeast GA Health System
2016	Kevin Lunceford	Pharmacist, Pruitt Health
2017	Aaron Chase	Pharmacist Resident, Augusta
2017	Sarah Payne	Pharmacist Resident, Mission Health
2018	Minna Hassan	PharmD Student, UGA
2018	Laura Pyronneau	Pharmacist, Kroger
2018	Catherine Rothery	Pharmacist Resident, Banner Health

Years	High School Student
2012	Ian Dawkins
2013-2014	Augustine Song
2015	Mary Glassman

PROFESSIONAL SERVICE

Department

2022-Present	Interim Department Head
2021-2022	Interim Associate Department Head
2020-2021	Interim Assistant Department Head
2021-Present	Member, Dept Head Search Committee
2019-Present	Chair, Faculty Mentoring Committee
2019-Present	Member, Executive Committee

2018-2019	Member, Committee for Research
2018-2019	Member, Faculty Search Committee, UGA Chemistry (Organic)
2018-2019	Member, Faculty Search Committee, PBS and CTEDG
2017-2018	Member, Faculty Search Committee, PBS
2014-2017	Chair, Seminar Committee
2015	Member, PBS Self-Study Committee
2014-2017	Executive Committee Member
2014-Present	Chair, Seminar Committee
2012-Present	Member, Chu Lectureship Planning Committee
2012-Present	Chair, Equipment Maintenance and Training
2011-2013	Member, Seminar Committee
2011-2013	Member, Social and Entertainment Committee
2011-2012	Member, Strategic Plan Task Force

College of Pharmacy

2022-Present	Administrative Committee
2021-2022	Chair, Faculty and Staff Awards Committee
2020-2021	Member, Faculty Council
2017-2019	Member, Graduate Education and Curriculum Committee
2016-2019	Member, Academic and Professionalism Committee
2014-2016	Chair, Computer and Instructional Technology Committee
2013-2014	Member, Computer and Instructional Technology Committee
2012-2013	Chair, Graduate Education and Curriculum Committee
2012	Chair, Ad hoc PharmD/PhD Program Committee
2011-2012	Committee Member, Graduate Education and Curriculum Committee

University

2022-Present	Director, UGA Cancer Center
2021	Member, UGA Pew Biomedical Scholar Award Committee
2021	Member, UGA Cancer Center Review Committee
2020-Present	Member, Campus-Wide Faculty Search Committee for President's Cluster Hiring Initiative
2020-2021	Member, International Research Collaborations Working Group
2019-Present	Member, UGA innovation District Faculty Advisory Group
2018-2019	Member, Faculty Search Committee (Dept. of Chemistry)
2017-2018	Member, Search Committee for COP Dean Position
2015	Invited Member, Faculty Focus Group, UGA OVPR eResearch Portal
2014-2022	Executive Committee Member, UGA Cancer Center
2012-2017	Executive Committee Member, Chemical Biology Group, Integrated Life Sciences Program
2010-Present	Member, Center for Drug Discovery
2010-Present	Member, Center for Undergraduate Research Opportunities
2011-Present	Member, Louis Stokes Alliance for Minority Participation
2011-Present	Member, Interdisciplinary Life Sciences
2011-Present	Member, Biomedical Health Sciences Institute

National

2020-2021	Chair, Young Investigator Award Committee, American Peptide Society (APS)
2019-2025	Elected Council Member, American Peptide Society (APS)

2019-2025	Member, International Liaison Committee, American Peptide Society (APS)
2019-Present	Member, SRC Advisory Committee, FASEB
2017-2019	Co-Chair, Student Affairs Committee, American Peptide Society (APS)
2015-2017	Elected Member, Nominating Committee, American Peptide Society (APS)
2012-2015	Committee Member, Mentoring and Career Development, ASPET
2011-2012	Committee Member, Diversity, ASPET

Service for Other Universities

2020	External Reviewer, BS, MS, and PhD Programs, Pharmaceutical Sciences, U. of South Carolina College of Pharmacy

EDITORSHIPS/EDITORIAL BOARD MEMBERSHIPS

2021-2022	Guest Editor, Special Issue on "Macrocyclic Peptides" for *Frontiers in Molecular Biosciences*
2018	Guest Editor, Special Symposium-in-Print issue "Constrained Peptides and Biological Targets" for *Bioorganic and Medicinal Chemistry,* March 2018
2015-Present	Review Board Editor, *Frontiers in Pharmacology*

GRANT REVIEWS/STUDY SECTIONS

2021	Member, ZRG1 BCMB-G 55 study section (NIH), October
2021	Ad hoc member, Synthetic Biological Chemistry B (NIH SBCB), June
2020	Member, ZRG1 BCMB-G 55 study section (NIH), October
2020	National Science Center (Poland)
2020	Wellcome Trust UK (United Kingdom)
2020	Michael J. Fox Foundation
2019	Ad hoc member, Synthetic Biological Chemistry B (NIH SBCB), October
2019	Cystic Fibrosis Canada
2019	Michael J. Fox Foundation, Staff-Initiated Projects
2018	Inserm Plan Cancer Programmes (France), EVA3 Grants
2017	Inserm Plan Cancer Programmes (France), Biologie des Systemes

GRANTS RECEIVED, 2010-Present:

1. **Michael J. Fox Research Grant**
   Time Period: 8/1/2020-7/31/2022
   Total Amount: $1,458,265
   Role: PI

2. **NIH R01 GM134097**
   Targeting WASF3 in tumor metastasis
   Time Period: 8/5/2019-7/31/2023
   Total Amount: $1,197,785
   Role: PI

3. **Michael J. Fox Idea Award**
   Time Period: 10/15/2018-2/15/2021
   Total Amount: $186,238
   Role: PI

4. **NIH P01 AG036675**
   Age Induced Impairment of Nutrient Signaling Results in Bone Loss
   Time Period: 04/01/2017– 03/31/2022
   Total Amount: $ 2,277,404 (Kennedy: $42,726)
   Role: Collaborator

5. **DoD Cancer Research Program: Idea Award**
   Therapy for the Adolescent/Young Adult Cancer Fibrolamellar Hepatocellular Carcinoma
   Time Period: 04/15/2017-3/15/2020 (NCE)
   Total Amount: $140,000
   Role: Co-Investigator

6. **COP Foundation Funds**
   AKAP Signaling in Liver Cancer
   Time Period: 2/21/2017-6/30/2018
   Total Amount: $5000
   Role: PI

7. **NIH R03 CA188439**
   Targeted inhibition of EGFR dimerization
   Time Period: 07/01/2015– 06/31/2017
   Total Amount: $ 150,000
   Role: PI

8. **NIH R01 EB016100**
   Secretory phospholipases $SPLA_2$ and their receptors for delivering nanoparticles
   Time Period: 1/1/2013-12/31/2016
   Total Amount: $1,345,892 (Kennedy: $30,000)
   Role: Co-Investigator (5%) (PIs: B.S. Cummings and R.D. Arnold)

9. **NCI Transition Career Development Award K22 CA154600**
   Probing the role of AKAPs in breast cancer using stapled peptide inhibitors
   Time Period: 9/01/11-8/31/15
   Total Amount: $571,000
   Role: PI

10. **COP Foundation Funds**
    Synthetic modulation of AKAP signaling
    Time Period: 10/08/2015-6/30/2016
    Total Amount: $3000
    Role: PI

11. **UGA-GRU Offices of the Vice President for Research**
    Targeting the WASF3/NCKAP1 interaction to suppress cancer cell invasion and metastasis
    Time Period: 9/1/2015-6/31/2016
    Total Amount: $37,500
    Role: PI

12. **UGA-GRU Cancer Centers**
    Targeting WAVE3 in tumor metastasis
    Time Period: 9/1/2014-8/31/2015

Total Amount: $37,500
Role: PI

13. **COP Foundation Funds**
Dissecting AKAP complexes
Time Period: 9/01/2014-6/30/2015
Total Amount: $3500
Role: PI

14. **Faculty Summer Research Support, Office of the Provost, UGA**
Time Period: 7/1/2013-6/30/2014
Total Amount: $5000
Role: PI

15. **UGA Pharmacy Special Project Fund**
EGFR Inhibition in Asthma
Time Period: 12/01/2010-06/30/2012
Total Amount: $22,000
Role: PI

16. **UGA College of Pharmacy Foundation Funds**
Uncovering the Role of AKIP1 Regulation on Androgen Receptor Localization
Time Period: 10/01/2011-06/30/2012
Total Amount: $4,000
Role: PI

17. **UGA College of Pharmacy Foundation Funds**
Targeting the EGFR dimerization interface as a novel strategy for inhibition.
Time Period: 11/01/2012-10/31/2013
Total Amount: $4,000
Role: PI